Republic of Iraq Ministry of Higher Education and Scientific Research University of Diyala College of Medicine



Molecular and Bacterial Study of *Helicobacter*pylori in Gastric Ulcer Patients in Baqubah Teaching Hospital

A Thesis

Submitted to Council College of Medicine - University of Diyala in
Partial Fulfillment of the Requirements for the Master Degree of
Science in Medical Microbiology

By

Nadia Hameed Jameel

BSc. Biology - University of Diyala

Supervised by

Lecturer Doctor

Assistant Professor

Dr. Anfal Shakir Motib

Dr. Ahmed Methab Athab

(Ph.D. in Molecular biology)

(MBChB-FICMS)

2020 A.D.

1441 A.H.

بسم الله الرحمن الرحيم

[نَرْفَعُ دَرَجَاتٍ مَّن نَّشنَاءُ وَفَوْقَ كُلٍ ذْي عِلْمِ عَلِيمٌ]

صدق الله العظيم

سورة يوسف {76}

Dedication

 Σ To the dearest people in my life "father & mother"

For their endless love, support and encouragement.

 Σ To the sisters and my husband.

 Σ To my friends with whom my life shines brightly.

 Σ To everyone who helped and supported me in my study.

NADIA

Acknowledgments

In the name of Allah, the most gracious, the most merciful, all praises be due to Allah, the sustainer of the entire world, the origin of science and wisdom, and may Allah's mercy and peace be upon our prophet, Mohammad, his relatives and companions.

I would like to express my sincere gratitude to my supervisor Dr. Anfal Shaker Motib and Dr. Ahmed Methab Athab for their continuous support, patience, motivation, enthusiasm, and immense knowledge.

I would like to thank the doctors in the endoscopic unit Dr. Abdelmohsin Razi Sadek &Dr. Sajed Ali Hussien, specialists in general surgery. And thank you to all members of endoscopic unit in Baqubah Teaching Hospital. Very special thanks to Dr. Walied Rahiem Al-Mineae in the staff of histopathology. Special thanks to Assistant prof. Burooj Mohameed Razooqi.

Special thanks are extended to staff of microbiology unit at Baqubah Teaching Hospital.

Finally, great thanks, deep appreciation and gratitude to my lovely family for their support and encouragement for me, and special thanks to my friends Sarah Ali Dawood & Nayarah Samer Hussien.

NADIA

Supervisor Certification

We, Certify that this thesis entitled (Molecular and Bacterial Study of Helicobacter pylori in Gastric Ulcer Patients in Baqubah Teaching Hospital) has been conducted under our supervision at College of Medicine, University of Diyala, as a partial requirements for the Master Degree of Science in Medical Microbiology.

Lecturer Doctor

Assistant Professor

Dr. Anfal Shakir Motib

Dr. Ahmed Methab Athab

In view of available recommendation, I forward this thesis for debate by the examining committee.

Signature

Assistant Professor Dr. Walaa Najm Abood

Head of Microbiology Department

College of Medicine – University of Diyala

Examination Committee Certification

We, the examining committee, certify that we have read this thesis entitled (Molecular and Bacterial Study of Helicobacter pylori in Gastric Ulcer Patients in Baqubah Teaching Hosbital) which prepared by (Nadia Hameed Jameel Al-Jobori) and have examined the student in its contents, and that in our opinion it is adequate for awarding the Degree of Master of science in Medical Microbiology.

Signature:

Name: Dr. Rajwa H. Essa

Scientific Degree: Professor

(Chairman)

Date: / /2020

Signature:

Signature:

Name: Dr.Adawia F. Abbas Name: Dr.Ali M. Ja'afer

Scientific Degree: Assist. Professor Scientific Degree: Assist. Professor

(Member) (Member)

Date: / /2020 Date: / /2020

Signature: Signature:

Name: Dr. Anfal SH. Motib Name: Ahmed M. Athab

Scientific Degree:Lecturer Doctor Scientific Degree:Assist.Professor

(Member/Supervisor) (Member/Co-Supervisor)

Date: / /2020 Date: / /2020

Approved by the Council of the College of Medicine – University of Diyala.

Signature:

Professor Dr.Ismail A.Latif

Dean

College of Medicine-University of Diyala

Date: / /2020

Summary

Helicobacter pylori is the most common causes of peptic ulcer disease, and it is one of causative agents of vitamin B12 deficiency. The infection by *H. pylori* transmitted by oral-oral, fecal-oral and gastric oral routs. The prevalence of this bacteria depend on age, sex, smoking or nonsmoking and also chronic disease such as diabetes.

The present study was conducted to determine the *H. pylori* infection and it is associated with cobalamin deficiency in patients with gastric ulcers which were diagnosed by using five different tests including, bacterial culture, rapid diagnostic test, rapid urease test, histopathology test and molecular test by detection 16SrRNA and some virulence factors like (UreA and CagA).

The study sample was 200, 127 (63.5%) males and 73 (36.5%) females age ranged from (10 to \geq 60) years were collected from Baqubah teaching hospitals, during the period from September 2018 till January 2019. From 200 individual 110 considered patients after initial diagnosis by endoscopic unit (presence symptoms) and 90 were considered control group (absence symptoms). Two types of samples were collected from each individual including gastric biopsy specimens for culture test, rapid urease test, histopathology test and polymerase chain reaction (PCR) to detect *H. pylori* infection, and blood samples were used for rapid diagnostic test to detect IgG antibodies of *H. pylori* and ELISA test to detect vitamin B12 deficiency.

The results showed that the presence of *H. pylori* in 3 patients was positive for bacterial culture, from these patients one patient appeared resistant to the antibiotic (amoxicillin, clarithromycin, tetracycline, metronidazole, ciprofloxacin, levofloxacin) but two patients showed resistance to some of these antibiotics used and sensitive to others.

In rapid diagnostic test the positive results was (109) (99.1 %), In rapid urease test the positive result from 110 patients was (86) (78.2 %), the positive results of histopathology test was 10 (9.0%), in molecular test genomic DNA was also extracted from gastric biopsies of all 200 individuals and used directly for PCR to detect *H. pylori* using 515 bp domain of (16SrRNA gene), which showed positive in 106 (96.4%) patients, and in virulence genes (UreA and CagA), the positive result of UreA 81 (73.60%) and CagA was 20 (18.20%). In addition, the extracted DNA of biopsies samples were sent for sequencing to identify the *H. pylori* strain. The result appeared that the strain of *H. pylori* that cause gastric ulcer in Baqubah city is *H. pylori* F211 which have the point mutation in 16SrRNA in cancer patients.

The incidence of infection in male was more than in female as the percentage was (70.00%) and (30.00%), respectively. Minimum age was 10 years and maximum was ≥ 60 . The highest age specific frequency in the individuals is in the age group was (40-49)&(50-59) years old. The incidence of H. pylori in smoking patients was 34 (30.90%), while it was 76 (69.10%) among non-smokers. In chronic diseases like diabetes the rate of infection was 59 (54.10%), hypertension in patients was percentage (1.80%), patients suffering from diabetes and hypertension was percentage (15.60%), patient with asthma and allergy (1.80%). While the rate of people who did not suffer from chronic diseases and those infected with H. pylori was (26.60%). The association between vitamin B12 and H. pylori showed that people infected with this bacterium suffered from vitamin B12 deficiency. The results showed that 110 patients with H. pylori infection have B12 deficiency.

The present study concluded that the PCR technique is the best method for the detection of these bacteria directly from gastric biopsy specimens, and vitamin B12 deficiency occur in patients infected with *H. pylori*.

Table of Contents

	Contents Page No.	
Dedicati		
	Acknowledgment	
Abstract		I
Table of contents		III
List of ta		VIII
List of fi	bbreviations	IX
List of a		X
	Chapter One	
1.1	Introduction	1
1.2	Aims of the study	4
	Chapter Two	
2	Literature Review	5
2.1	General description of <i>Helicobacter</i> pylori	5
2.2	Prevalence of <i>H. pylori</i> infection	6
2.3	Clinical feature	7
2.4	Epidemiology	8
2.5	Pathogenesis	9
2.6	Sources and transmission of <i>H. pylori</i>	10
2.7	Diagnosis of <i>H. pylori</i> infection	11
2.7.1	Invasive tests	11
2.7.1.1	Endoscopy	11
2.7.1.2	Histology	12
2.7.1.3	Rapid Urease Test	13
2.7.1.4	Bacterial culture	13
2.7.1.5	Polymerase chain reactions (PCR)	13
2.7.2	Non-invasive tests	14

2.7.2.1	Urea breath test (UBT)	14
2.7.2.2	Serological IgG test	15
2.7.2.3	H. pylori stool antigen testing	15
2.8	Treatment	16
2.9	Antibiotic resistance	17
2.10	Virulence factors	17
2.11	Environmental factors are involved in virulence	21
2.11.1	Gut flora	22
2.11.2	Smoking	23
2.11.3	High salt intake	23
2.11.4	Iron levels	23
2.11.5	Proton pump inhibitors (PPIs)	24
2.11.6	Di (2-ethylhexyl) phthalate	24
2.11.7	Vitamin B12	24
2.12	Association between <i>H. pylori</i> and vitamin B12 deficiency	26
	Chapter Three	
3	Patients, Materials and Methods	27
3.1	Patient and sample collection	27
3.1.1	Biopsy Samples	27
3.1.2	Blood Samples	27
3.2	Materials	28
3.2.1	Laboratory equipment's	28
3.2.2	Tools	29
3.2.3	Cultures Media	30
3.2.4	Cultures Wedia	
	The Diagnostic Kits	30
3.2.5		
3.2.5	The Diagnostic Kits	30
	The Diagnostic Kits The Stains	30
3.2.6	The Diagnostic Kits The Stains Chemical materials	30 31 31

3.2.8.1	Oxidase Reagent	32
3.2.8.2	Catalase Reagent	32
3.2.8.3	Skirrows Solution	32
3.2.8.4	Preparation Tris-Borate buffer	33
3.2.8.5	Ethidium Bromide	33
3.2.8.6	Human Blood	33
3.2.8.7	Normal saline	33
3.2.8.8	Red safe staining souluion	34
3.2.9	Cultures medium	34
3.2.9.1	Skirrow's medium	34
3.2.9.2	Blood Agar	35
3.2.9.3	Brain Heart Infusion Broth	35
3.3	Methods	36
3.3.1	Culture of samples	36
3.3.2	Rapid urease test	36
3.3.3	H. pylori diagnosis and sensitivity	36
3.3.3.1	Gram stain	37
3.3.3.2	Oxidase test	37
3.3.3.3	Catalase test	37
3.3.3.4	Urease test	37
3.3.3.5	Antibiotic Susceptibility Test	37
3.3.4	Test detection about IgG for <i>H. pylori</i> by use One Step <i>H. pylori</i> Test Device Kit	38
3.4	Histopathologic Examination	39
3.5	Molecular methods for <i>H. pylori</i> detection	39
3.5.1	DNA Extraction	39
3.5.1.1	Extraction of genomic DNA from tissue biopsy	39
3.5.2	Agarose Gel Electrophoresis	41
3.5.2.1	Preparation of Agarose gel	41
3.5.2.2	Preparation of sample(loading DNA in agarose gel)	42
3.5.2.3	Primers Preparation	42

3.5.2.4	Preparation of polymerase chain reaction	42
3.5.2.5	The PCR amplification of 16SrRNA for H. pylori	43
3.5.2.6	The PCR amplification of UreA for H. pylori	44
3.5.2.7	The PCR amplification of CagA for H. pylori	44
3.5.3	Detection of the product of PCR and molecular size estimation	45
3.6	Gel Extraction (Sequencing) Protocol	45
3.6.1	DNA Sequencing	46
3.7	Serology Test	47
3.7.1	Enzyme-Linked Immunosorbent Assay Testing (ELISA)	47
3.8	Statistical analysis	48
	Chapter Four	
4	Results	49
4.1	Sample description	49
4.2	Results of diagnostic methods	51
4.2.1	Bacteriology culture test	51
4.2.2	antibiotic resistance	52
4.2.3	Rapid diagnostic test(RDT)	52
4.2.4	Rapid urease test	54
4.2.5	Histopathology results	55
4.3	Molecular methods for <i>H. pylori</i> detection	56
4.3.1	Detection of 16SrRNA gene of <i>H. pylori</i>	56
4.3.2	Determination of <i>Helicobacter pylori</i> Virulence Genes by PCR	57
4.3.3	DNA sequencing	60
4.3.4	Tree analysis of DNA	62
4.4	The evaluation of diagnostic methods for <i>H. pylori</i> detection	63
4.5	The incidence of <i>H. pylori</i> among individuals in relation to variable factors	64
4.5.1	The distribution of <i>H. pylori</i> infection among individuals according to sex	64
4.5.2	The distribution of <i>H. pylori</i> infection among individuals according to age groups	64
`4.5.3	The distribution of <i>H. pylori</i> among individuals according to smoking	65

4.5.4	The relationship between chronic diseases and <i>H. pylori</i> infection	66
4.5.5	The distribution of <i>H. pylori</i> infection according to the cancer	67
4.5.6	ELISA test for measuring VB12	68
	Chapter Five	
5	Discussion	69
5.1	sample collection	69
5.2	The laboratory diagnosis	70
5.2.1	Bacteriology culture test and antibiotic sensitivity	70
5.2.2	Rapid diagnostic test(RDT)	71
5.2.3	Rapid urease test(RUT)	71
5.2.4	Histopathology test	72
5.2.5	Pcr techniques	73
5.2.6	DNA sequencing	75
5.3	The evaluation of diagnostic methods for <i>H. pylori</i> detection	75
5.4	The distribution of <i>H. pylori</i> among individuals according to sex and age groups	76
5.5	The distribution of <i>H. pylori</i> among individuals according to smoking	77
5.6	The relationship between chronic diseases and <i>H. pylori</i> infection	78
5.7	The distribution of <i>H. pylori</i> infection according to cancer	79
5.8	ELISA test for detection VB12	79
	Chapter Six	
6.1	Conclusions	81
6.2	Recommendations	81
	References	
	References	82-106
	Appendix	
	Appendix(1)	
5.5 5.6 5.7 5.8	according to sex and age groups The distribution of <i>H. pylori</i> among individuals according to smoking The relationship between chronic diseases and <i>H. pylori</i> infection The distribution of <i>H. pylori</i> infection according to cancer ELISA test for detection VB12 Chapter Six Conclusions References References Appendix	77 78 79 79 81 81

List of Tables

Table	Title	Page
No.		No.
3-1	The general equipment's used in this study	28
3-2	The general tools used in the present study	29
3-3	Cultures media used in this study	30
3-4	The diagnostic kits are used in this study	30
3-5	The stains are used in this study	31
3-6	The chemical materials used in this study	31
3-7	The primers used in present study	32
3-8	The contents of DNA extraction kit	39
3-9	Components of the reaction medium used for DNA replication	39
3-10	PCR conditions for 16SrRNA gene primer	43
3-11	PCR conditions for UreA primer	44
3-12	PCR conditions for CagA primer	44
4-1	Results bacteriology culture test of biopsy tissues	51
4-2	The resistance of <i>H. pylori</i> isolates for antibiotics	52
4-3	The results of Rapid diagnostic test	53
4-4	The results of Rapid urease test	54
4-5	Histopathology test results with patients groups	55
4-6	The percentage of 16SrRNA according to individuals	56
4-7	Comparative positivity tests specific for <i>H. pylori</i> infection	63
4-8	The frequency of <i>H. pylori</i> infection among individuals according to sex	64
4-9	The frequency of <i>H. pylori</i> infection among individuals according to age groups	65
4-10	The frequency of <i>H. pylori</i> infection among individuals according to chronic diseases	67

List of Figures

Figure	T:41-	Page
No.	Title	No.
2.1	Schematic representation of selected virulence factors in <i>Helicobacter pylori</i>	21
4-1	The colonies of <i>H. pylori</i> on skirrows medium	51
4-2	Rapid diagnostic test (RDT) for detection of <i>H. pylori</i>	53
4-3	Rapid urease test result	54
4-4	The results of histopathology test	55
4-5	Agarose gel electrophoresis of DNA from biopsy tissue directly showing PCR products for 515 bp of 16SrRNA	57
4-6	Agarose gel electrophoresis of DNA from biopsy tissue directly showing PCR products for 411bp of UreA	58
4-7	The percentage of present UreA gene in patients infected with <i>H. pylori</i>	58
4-8	Agarose gel electrophoresis of DNA from biopsy tissue directly showing PCR products for 1320bp of CagA	59
4-9	The percentage of CagA gene in patients infected with <i>H. pylori</i>	59
4-10	Alignment statistics for DNA sequencing. The results showed that the strain is <i>H. pylori</i> strain F211 and the identity between them is 100%	60
4-11	Alignment statistics for DNA sequencing of cancer patients. The results showed that the DNA of <i>H. pylori</i> strain F211 but they have transvertion mutation and the identity between them is 99%.	61
4-12	Tree analysis for DNA samples which were extracted from biopsies patients suffering from gastric ulcer	62
4-13	The distribution of <i>H. pylori</i> infection among individuals according to the smoking	66
4-14	The distribution of <i>H. pylori</i> infection in individuals according to cancer	67
4-15	Vitamin B12 level in the individuals	68

List of Abbreviations

Abbreviate	Key
Abs	Antibodies
Ag	Antigen
CagA	Cytotoxin associated gene activity
BabA	Blood group antigen binding Adhesion
EDTA	Ethyle Dimethyl Tetra Acitc acid
PCR	Polymerase Chain Reaction
OMP	Outer Membrane Protein
MALT	Mucosa-Associated Lymphoid Tissue
LPS	Lipoplysaccharide
P.P.I	Proton Pump Inhibitor
рН	Potential hydrogen
rRNA	ribosomal Ribonucleic acid
S	Svedberg
UBT	Urea Breath Test
VacA	Vacuolating cytotoxin activity
WHO	World Health Organization
If	Intrinsic factor
ELISA	Enzyme linked immunosorbent assay
DEHP	Di (2-ethylhexyl) phthalate

Chapter One Introduction

1.1 Introduction

Helicobacter pylori is a gram-negative bacterium and spiral in shape, which colonizes the human stomach mucoid lining (Smyk et al., 2014). It is characterized by polymorphism phenomenon and it may appear as coccoid and bacillary form (Mamoun et al., 2015). It is the main cause of stomach and duodenal ulcers, which have become common in recent times due to the spread this type of bacteria, that are highly pathogenic, affect more than half of the world population (Nevine et al., 2015; Mamoun et al., 2015).

The incidence of this bacterium is due to the virulence genes (cag A and vacA) that are carried by particular genetic patterns of *H. pylori*, which are the most important virulence genes accompanying stomach and bowel disease (Salimzadeh *et al.*, 2015; Wang *et al.*, 2015).

To avoid the harsh condition in the gastric lumen, this bacterium has developed resistance to the stomach acid through colonization in a very narrow place of gastric lactation and secretion of the urease which break down urea located in the medium to ammonia which have the effect of the acidic acid around in the stomach lining which enables them to stay in the human stomach lifelong if not treated with antibiotics (Bakir *et al.*, 2012). Therefore, this bacterium causes many diseases such as chronic gastritis, gastric ulcers, duodenal ulcers, gastric cancer and mucosa-associated lymphoid-tissue lymphoma (Erzooki *et al.*, 2016).

The transmission of *H. pylori* may occur via oral-oral, fecal-oral, gastric-oral or iatrogenic routes (Song *et al.*, 2000). The prevalence of *H. pylori* infection varies widely according to the age, sex, race and ethnicity (Jackson *et al.*, 2009).

A number of studies have shown the highest rates of infection are associated with low socioeconomic status, family size, crowding, low level of education, poor sanitation and uncleanly water supplies (Al-Sulami *et al.*, 2012).

Diagnosis of *H. pylori* infection can be made by using several invasive or non-invasive techniques. Invasive diagnostic assays include: rapid urease test, histological examination, culture and polymerase chain reaction (PCR). Non-invasive diagnostic assays include serology tests, urea breath test and stool antigen (Behnam *et al.*, 2015).

These bacteria are characterized as being fastidious, they appear very poorly in the tissue, making it difficult to develop and considered a slow micro- organism and therefore proposes to be diagnosed directly from clinical models by using molecular techniques like (PCR) which is unique in its sensitivity and high specificity in diagnosis and accuracy in determining both the presence of infection and the genotype for these bacteria (Abu-Sbeih *et al.*, 2014).

Some studies have shown that culture assay is used as a gold standard to detect patients with active H. pylori infection in clinical samples (Al-Jobori et al., 2011; Ramis et al., 2012). A study in Iraq by Bakir et al. that collected 92 gastric biopsy samples, 25 H. pylori negative and 67 H. pylori positive patients. In H. pylori positive group, the positive rates of H. pylori DNA extraction in the gastric epithelial cells were increased in chronic superficial gastritis, precancerous changes and gastric cancer groups (P>0.01) (Bakir et al., 2012). In another study in Thi-Qar, were collected from 70 patients (35 male and 35 female). The results showed that there is a significant increasing (p \leq 0.05) in H. pylori in males (71.43 %) when compare with females patient

(24.59%) and negative result in males patient (28.57%) while the negative result in females patient are (45.71%) (Hussein *et al.*, 2015).

The detection of (CagA) in positive *H. pylori* is higher in patients who had gastric cancer compare to those with chronic superficial gastritis and atrophic gastritis (P<0.01) (Bakir *et al.*, 2012). *Helicobacter pylori*-specific region of 16SrRNA sequence is high conserve among most *H. pylori* strains and allow specific detection and identification of this bacterium in biological specimens (Liu *et al.*, 2008).

H. pylori infection has a strong association with chronic infection of the stomach, that lead to impairment in the gastric acid and pepsin production, which lead to impair absorption of food and vitamin B12. Therefore, this bacterium may cause a cobalamin deficiency and it is a risk factor for gastritis ulcers (Kadhim *et al.*, 2015).

1.2 Aims of the study

The main aims of this study are to:

1- Isolation and identification of *H. pylori* from biopsies samples of patients suffering from gastric ulcer and determine the antibiotic resistance of *H. pylori*.

- 2- Detection of *H. pylori* by using:
 - A) Rapid diagnostic test in serum patients.
 - B) Rapid urease test and histopathology test in gastric biopsies samples.
 - C) PCR of 16SrRNA.
- 3- Finding the correlation with different parameters such as gender, age, smoking, chronic disease and cancer.
- 4- Evaluate the rate of gastric ulcer infection with *H. pylori* and its relation with vitamin B12 deficiency.