

Antimicrobial Susceptibility Patterns of Enterococcal Isolates And its Relevance With Biofilms Formation And B -Lactamase Production

*Abdul-Razak SH. Hasan (Ph.D.) **Abbas A. Al-Duliami (Ph.D.) ***Kariem S. Al-Ajeely (Ph.D.) ****Zainab H. Al-Azawi (M.Sc.) Diyala, Iraq. Abstract

Background: Enteroccci are part of the normal intestinal flora of human and animal, but with increasing antimicrobial resistance, enterococci are recognized as serious nosocomial as well as community pathogens.

Objectives: To investigate the antimicrobial susceptibility patterns of 44 isolates of enterococci recovered from different pathological specimens from in-and out-patients from Diyala province.

Materials and methods: The present study was conducted in Baquba General Hospital and Al-Batool Hospital for Maternity and children during the period from 1st. September/2005 to 30th. September /2006. A total of 343 specimens were collected from 213 inpatients and 130 outpatients. 200 (58.3%) were females and 143 (41.7%) were males. The mean age of patients was (32.8 ± 17.2) years. Specimens include, urine, stool, vaginal swabs, throat swabs, burn swabs, blood for culture, middle ear swabs, wound swabs, sputum and cerebrospinal fluid. Specimens were streaked on blood agar, and other differential and selective media. 44 isolates of enterococci (30 E. faecalis, 10 E. faecium, 3 E. gallinarium, and 1 E. avium) were recovered and identified according to standard bacteriological and biochemical criteria. The susceptibility patterns toward 13 antimicrobial agents were done by disc diffusion method. Data were statistically analysed.

Results: The results revealed that the highest susceptibility of enterococcal isolates was toward the Nalidixic acid (79.5%), Ciprofloxacin (61.4%), Amoxacillin+clavilanic acid (61.4%), Rifampicillin (36.4%), Trimethoprim (22.7%), Vancomycin (11.4%). However, all isolates were resistant to Cloxacillin, Cefotaxim, Amoxicillin, Tetracycline, and Erythromycin. The susceptibility of non- β -lactamase producing isolates to penicillin were significantly higher than β - lactamase producing isolates (p<0.001). Furthermore, the sensitivity of non-biofilms former isolates were significantly higher than that of biofilms former isolates (p=0.002).

Conclusion: The overall susceptibility rates of enterococcal isolates recovered from nosocomial as well as community acquired infections to available antimicrobials are low. **Keywords:** Enterococci, E. faecalis, Antimicrobial susceptibility.

*College of Medicine- Diyala University **College of Education- Dyala University ***Colege of Vet. Medicine-Diyala University ****College of Education- Diyala University

Diyala, Iraq.

Introduction

Enterococci are widely distributed in the environment, they are normal commensals that usually inhabit the alimentary tract of human in addition to being isolated from environmental and animal sources [1,2]. E. faecalis and E. faecium are the most prevalent species cultured from human, accounting for more than 90% of clinical isolates. Other enterococci species to cause human infection include, E. avium, E. gallinarum, E. casseliflavus, E. durans, E. raffinosus, and E. mundtii [3-5].

In the past 15 years, enterococci have emerged as increasingly important cause of acquired nosocomial infections worldwide, including urinary tract infection, bacteremia, surgical wound infection, intraabdominal and pelvic infection, endocarditis, and meningitis [6-9]. An alarming fact is the intrinsic resistance to many antimicrobial agents and acquisition of resistance to other the antibiotics available for treatment has led to [10-12] therapeutic difficulties worldwide Studies on the antimicrobial susceptibility patterns of enterococci have affirmed the worldwide emergence of multiple-drug enterococci. particularly resistant vancomycin [13-17].

It has been documented that nosocomial enterococci have numerous virulence factors that enhance their ability to colonize hospitalized patients, contribute to antimicrobial resistance, and aggravate the outcome [18-20]. Among the virulence factors are biofilms formation and ßlactamase production. It has been reported that biofilms formation capacity is restricted to enterococci harboring enterococci surface protein which promotes primary attachment and biofilms formation [21,22]. On the other hand, β-lactamase producing enterococci have acquired resistance to penicillins,

ephalosporins, carbapenems a monobactams [23,24].

Materials and methods

The present study was conducted in Baquba General Hospital and Al-Batool Hospital for Maternity and children during the period from 1st. September/2005 to 30th. September /2006. A total of 343 specimens were collected from 213 inpatients and 130 outpatients. 200 (58.3%) were females and 143 (41.7%) were males. The mean age of patients was (32.8 ± 17.2) years. Specimens include, urine, stool, vaginal swabs, throat swabs, burn swabs, blood for culture, middle ear swabs, wound swabs, sputum and cerebrospinal fluid. Specimens were streaked on blood agar, and other differential and selective media. 44 isolates of enterococci (30 E. faecalis, 10 E. faecium, 3 E. gallinarium, and 1 E. avium) were recovered identified according to standard and bacteriological and biochemical criteria. The ability of β -lactamase production was detected according to the method described by [25]. Detection of biofilms formation was followed the method of ^[26]. The susceptibility patterns toward 13 antimicrobial agents were disc done by diffusion method. Determination of sensitive resistant or antimicrobial was based National on Committee for Clinical Standards (NCCLS) [27]. Data were statistically analyzed.

Results

The results in table (1) revealed that the highest sensitivity rate of enterococci isolates was toward the Nalidixic acid (79.5%), followed by Ciprofloxacin, Amoxacillin+clavilanic acid (61.4%),Rifampicillin (36.4%), Trimethoprim (22.7%), vancomycin (11.4%). However, all isolates were resistant to Cloxacillin. Cefotaxim, Amoxicillin, Tetracycline, and Erythromycin.





	Enterococcal isolates				Total
Antimicrobiala	E. faecalis	E. faecium	E. gallinarium	E. avium	(n-44)
Anumicrobiais	(n=30)	(n=10)	(n=3)	(n=1)	(11-44)
	No. (%)	No. (%)	No. (%)	No. (%)	No. (%)
Vancomycin	0(0)	2(20)	3(100)	0(0)	5(11.4)
Ciprofloxacin	18(60)	7(70)	1(33.3)	1(100)	27(61.4)
Rifampicin	11(36.7)	5(50)	0(0)	0(0)	16(36.4)
Nalidixic acid	25(83.3)	7(70)	2(66.7)	1(100)	35(79.4)
Penicillin	7(23.3)	4(40)	0(0)	1(100)	12(27.3)
Amoxacillin +	16(53.3)	8(80)	3(100)	0(0)	27(61.4)
clavulanic acid			0		
Trimethoprim	6(20)	3(30)	1(33.3)	0(0)	10(22.7)

Table 1: Antimicrobial susceptibility of enterococcal isolates.

Table(2)showedtheantimicrobialsusceptibilitypatternsofE.faecalisincomparisonwithotherenterococcalisolates.AllisolatesofE.faecaliswereresistantto

vancomycin, while 5(35.7%) of other enterococcal isolates were sensitive to it. The difference between the two groups was statistically significant (p=0.002).

	Enterococc		
Antimicrobials	Other species (n=14)	E. faecalis (n=30)	P (Fisher's exact)
	No. (%)	No. (%)	
Vancomycin	5(35.7)	0(0)	0.002 [S]
Ciprofloxacin	9(64.3)	18(60)	1 [NS]
Rifampicin	5 (35.7)	11(36.7)	1 [NS]
Nalidixic acid	10 (71.4)	25 (83.3)	0.43 [NS]
Penicillin	5 (35.7)	7 (23.3)	0.43 [NS]
Amoxacillin C +	11 (78.6)	16 (53.3)	0.18 [NS]
clavulanic acid			S
Trimethoprim 💽	4(28.6)	6 (20)	0.7 [NS]

Table 2: Antimicrobial susceptibility according to enterococcal species.

Regarding the effect of β -lactamase production on susceptibility to antimicrobial agents, the results revealed that the susceptibility of non- β -lactamase producing isolates to penicillin were significantly higher than β - lactamase producing isolates (p<0.001). Additionally, the resistance of β - lactamase producing isolates to Vancomycin, Ciprofloxacin, Rifampicin Nalidixic acid and Trimethoprim were insignificantly higher than that of non β lactamase producing isolates. Moreover, the sensitivity of β - lactamase producing isolates to Amoxacillin + clavulanic acid was insignificantly higher than non β -lactamase producing isolates, table (3).



	β -lactamase p	D (Fisher's	
Antimicrobials	Non-producer (n=12)	producer (n=32)	P (FISHER'S
	No. (%)	No. (%)	exact)
Vancomycin	1(23.1)	4(12.5)	1 [NS]
Ciprofloxacin	8(66.7)	19(59.4)	0.74 [NS]
Rifampicin	5(41.7)	11(34.4)	0.73 [NS]
Nalidixic acid	10(83.3)	25(78.1)	1 [NS]
Penicillin	11(91.7)	1(3.1)	< 0.001 [S]
Amoxacillin + clavulanic acid	7(58.3)	20(62.5)	1 [NS]
Trimethoprim	3(25)	7(21.9)	1 [NS]

Table 3: Antimicrobial susceptibility according to β-lactamase production.

The results also showed that the sensitivity of non-biofilms former isolates were significantly higher than that of biofilms former isolates (p= 0.002). While the sensitivity to Amoxacillin + clavulanic acid was insignificantly higher in

non-biofilms former compared to biofilms former isolates. On the other hands, biofilms former isolates were insignificantly more resistant to Rifampicin and Nalidixic acid compared to non-biofilms former isolates.

Table 4: Antimicrobial susceptibility according to biofilms formation.

	Biofilms for		
Antimicrobials	Non-formers (n=10)	Formers (n=34)	P (Fisher's exact)
	No. (%)	No. (%)	
Vancomycin	0(0)	5 (14.7)	0.57 [<mark>N</mark> S]
Cipr <mark>ofl</mark> oxacin	6(60)	21(61.8)	1 [NS]
Rifampicin	4(40)	12 (35.3)	1 [NS]
Nalidixic acid	9 (90)	26 (76.5)	0.66 [NS]
Penicillin	7 (70)	5 (14.7)	0.002 [S]
Amoxacillin + clavulanic acid	7(70)	20 (58.8)	0.72 [NS]
Trimethoprim	2(20)	8 (23.5)	1 [NS]

Discussion

The results showed that 79.5% of all enterococcal isolates (100% E. faecalis, 80% E. faecium and 100% E. gallinarium) were resistant to vancomycin. These results are consistent with previous studies [15-17]. The vancomycin resistant enterococci (VRE) have caused hospital outbreaks worldwide, and the vancomycin resistant gene (vanA) has crossed genus boundaries to methicillin resistant Staphylococcus aureus (MRSA). Spread of VER therefore represents an immediate thread for patients care and creates a reservoir for mobile resistance genes for other, more virulent pathogens [28, 29]. The first VRE isolates that harbored the van A transposon were identified in 1987 in Europe, and within 10 years VRE

represented > 25% of enterococci associated with nosocomial bloodstream infections in USA [30]. Recently, vancomycin resistant rate among E. faecalis and E. faecium were 5.4% and 75.4% respectively in USA [31]. The acquisition of vancomycin resistance by enterococci has seriously affected the treatment and infection control of these organisms. VRE, particularly E. faecium isolates, are frequently resistant to all antibiotics that are effective in the treatment vancomycin-susceptible enterococci, of which leaves clinicians treating **VRE** infections with limited therapeutic options [10.32].

The β -lactamase producing enterococci have significantly higher rate for penicillin resistance compared to β -lactamase non-



producing isolates. Additionally, 87.5% of βproducing enterococci lactamase were resistant to vancomvcin. These results are not unusual and are in concordant with previous reports[23,24]. Moreover, high level gentamicin resistance documented was among β - lactamase producing E. faecalis that are strongly associated with patients of severe underlying diseases [32].

References

[1]Brooks, G.F.; Carroll, K.C.; Butel, J.S. and Morse, S.A. The streptococci. In: Medical Microbiology. 24th. Ed. 2007. McGraw Hill.233-49.

[2]Fisher, K. and Phillips, C. The ecology, epidemiology and virulence of enterococci. Microbiology 2009; 155(6): 1749-57.

[3]7. Udo, E.E.; Al-Sweih, N.; Phillip, Q.A. and Chugh, T.D. Species prevalence and antibacterial resistance of enterococci isolated in Kuwait hospitals. J.Med. Microbiol. 2003; 52: 163-8.

[4]Quinones, D.; Goni, P.; Rubio, M.; Duran, E. and Gomez-lus, R. Enterococci species isolated from Cuba: species frequency of occurrence and antimicrobial susceptibility profile. Diag. Microbiol. Infect. Dis. 2005; 51(1): 63-7.

[5]Hasan, A. SH.; Al-Duliami, A.A. and Al-Azawi, Z. H. Species prevalence of enterococci isolated from hospital and community-acquired infections in Diyala province. Diyala J. Appl. Res. 2009; 5(1):

[6] Al-Otaibi, F.E.; Kambal, A.M. and Baabbad, R.A. Enterococcal bacteremia in a teaching hospital in the central region of Saudi Arabia. Saudi Med. J. 2004; 25(1): 21-5.

[7]Hunt, C.P. The emergence of enterococci as a cause of nosocomial infection. Brit. J. Biomed. Sci. 1998; 55(2): 149-56.

[8]Babay, H.A.; Twum-Danso, K.; Kambal, A.M. and Al-Qtaibi, F.E. Blood stream infections in pediatric patients. Saudi Med. J. 2005; 26(10): 1555-61.

[9]Akkoyun, S.; Kuloglu. F. and Tokuc, B. Etiologic agents and risk factors in nosocomial urinary tract infections. Microbiol. Bul. 2008;42(2): 245-54. [10] Spera, R.V. and Farber, B.F. Multi-drug resistant Enterococcus faecium: An untreatable nosocomial pathogen. Drugs, 1994; 48(5): 678-88.

[11] Sood, S.; Malhotra, M.; Das, B.K. and Kapil, A. Enterococcal infections and

antimicrobial resistance. Indian J. Med. Res. 2008; 128(2): 111-21.

[12]Johnson, P.J.; Townsend, J.P.; Bohn, T.; Simonsen, G.S.; Sundsford, A. and Nielsen, K.M. Factors affecting the reversal of antimicrobial-drug resistance. Lancet Infect. Dis. 2009; 9(6): 357-64.

[13]Huycke, M.M.; Sahm, D.F. and Gilmore, M.S. Multiple-drug resistant enterococci: the nature of the problem and an agenda for the future. Emerg. Infect. Dis. 1998; 4(2): 239-49.

[14] Klibi, N.; Gharbi, S.; Masmoudi, A.; Ben Slama, K.; Poeta, P.; Zarazaga, M.; Fendri, C.; Boudabous, A. and Torres, C. Antibiotic resistance and mechanisms implicated in clinical enterococci in a Tunisian hospital. J. Chemother. 2006; 18(1): 20-6.

[15]Udo, E.E.; Al-Sweih, N.; John, P. and Chugh, T.D. Antibiotic resistance of enterococci isolated at a teaching hospital in Kuwait. Diag. Microbiol. Infect. Dis. 2002; 43(3): 233-8.

[16] Zhanel, G.G.; DeCorby, M.; Laing, N.; Weshnoweski, B.; Vashisht, R.; et al. Antimicrobial-resistant pathogens I intensive care units in Canada: results of the Canadian National Intensive Care Unit (CAN-ICU) study, 2005-2006. Antimicrob. Agents Chemother. 2008; 52(4): 1430-7.

[17]Werner. G.; Coque, T.M.; Hammerum, A.M.; Hope, R.; Hryniewicz, W.; et al. Emergence and spread of vancomycin resistance among enterococci in Europe. Euro. Surveill. 2008; 13(47): 19046.

[18]Klibi, N.; Ben Slama, K.; Saenz, Y.; Masmoudi, A.; Zanetti, S.; Sechi, L.; Boudabous, A. and Torres, C. Detection of virulence factors in high-level gentamicinresistant Enterococcus faecalis and Enterococcus Faecium isolates from a Tunisian hospital. Can. J. microbial. 2007; 53(3): 372-9.



[19]Dupont, H.; Vael, C.; Muller, C.; Chosidow, D.; Mantz, J.; et al. Prospective evaluation of virulence factors of enterococci isolated from patients with peritonitis: impact on outcome. Diag. Microbiol. Infect. Dis. 2008; 60(3):247-53.

[20]Hallgren, A.; Claesson, C.; Saeedi, B.; Monstein, H.; Hanberger, H. and Nilsson, L. Molecular detection of aggregation substance, enterococcal surface protein, and cytolysin genes and in vitro adhesion to urinary catheters of Enterococcus faecalis and Enterococcus faecium of clinical origin. Int. J. Med. Microbiol. 2009; 299(5): 323-32. [21] Toledo-Arana, A.; Valle, J.; Solano, C.; Arrizubieta, M.; Cucarella, C.; et al., The enterococcal surface protein, Esp, is involved in Enterococcus faecalis biofilm formation. Appl. Envirn. Microbiol. 2001; 67(10): 4538-45.

[22]Mohamed, J.A. and Huang, D.B. Biofilm formation by enterococci. J. Med. Microbiol. 2007; 56(12): 1581-8.

[23]Rice, L.B. and Murray, B.E. Betalactamase producing enterococci. Dev.Biol. Stand. 1995; 85:107-14.

[24]McAlister, T.; George, N.; Faoagali, J. and Bell, J. Isolation of beta-lactamase positive vancomycin resistant Enterococcus faecalis; first case in Australia. Commun. Dis. Intell. 1999; 23(9): 237-9.

[25]Koneman, E.W.; Allen, S.; Janda, W.;
Scheckenber, P. and Winn, J. Color plate and
Textbook for Diagnostic Microbiology.
4th.Ed. Lippincott company, Washington.(2007-2008). I
2009; 65(2): 15
[32]Linden, F
vancomycin-re
Drugs 2002; 62429-39.Dubble Company (Company)
Company (Company)Dubble Company (Company)
Company)

[26]Sandoe, J.A.; Witherden, I.R.; Cove, J.H.; Heritage, J. and Wilcox, M.H. Correlation between enterococcal biofilms formation in vitro and medical-device related infection in vivo. J. Med. Microbiol. 2003; 2: 547-50.

[27]NationalCommitteeforClinicalLaboratoryStandards(NCCLS).Performancestandards for antimicrobial discsusceptibilitytests.12th.Ed. pp 50-76.

[28]Willems, R.J.; Top, J.; Van Santen, M.; Robinson, A.; Coque, T.M.; et al. Global spread of vancomycin-resistant Enterococcus faecium from distinct nosocomial genetic complex. Emerg. Infect. Dis. 2005;11(6): 821-8.

[29] Bonten, M.J.; Willems, R. and Weinstein, R.A. Vancomycin resistant enterococci: Why are they here, and where do they come from?. Lancet Infec. Dis. 2001; 1(5): 314-25.

[30]Center for Disease Control and Prevention. National Nosocomial infections Surveillance (NNIS) system report, data summary from January 1992- June 2001, issued August 2001. Am. J. Infect. Control 2001; 29: 404-21.

[31]Sader, H.S. and Jones R.N. Antimicrobial susceptibility of Gram-positive bacteria isolated from US medical centers: results of Daptomycin Surveillance Program (2007-2008). Diag. Microbiol. Infect. Dis. 2009; 65(2): 158-62.

[32]Linden, P.K. Treatment options for vancomycin-resistant enterococcal infections. Drugs 2002; 62(3): 425-41.